



Draft Genome Sequences of Six Ruminant *Coxiella burnetii* Isolates of European Origin

Sidi-Boumedine, Karim; Ellis, Richard J.; Adam, Gilbert; Prigent, Myriam; Angen, Øystein; Aspán, Anna; Thiéry, Richard; Rousset, Elodie

Published in:
Genome Announcements

Link to article, DOI:
[10.1128/genomeA.00285-14](https://doi.org/10.1128/genomeA.00285-14)

Publication date:
2014

Document Version
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

Citation (APA):
Sidi-Boumedine, K., Ellis, R. J., Adam, G., Prigent, M., Angen, Ø., Aspán, A., Thiéry, R., & Rousset, E. (2014). Draft Genome Sequences of Six Ruminant *Coxiella burnetii* Isolates of European Origin. *Genome Announcements*, 2(3), [e00285-14]. <https://doi.org/10.1128/genomeA.00285-14>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Draft Genome Sequences of Six Ruminant *Coxiella burnetii* Isolates of European Origin

Karim Sidi-Boumedine,^a Richard J. Ellis,^b Gilbert Adam,^a Myriam Prigent,^a Øystein Angen,^{c*} Anna Aspán,^d Richard Thiéry,^a Elodie Rousset^a

ANSES Sophia-Antipolis Laboratory, Sophia-Antipolis, France^a; Animal Health and Veterinary Laboratories Agency, AHVLA Weybridge, New Haw, Surrey, United Kingdom^b; National Veterinary Institute, Frederiksberg C, Denmark^c; National Veterinary Institute of Sweden, Uppsala, Sweden^d

* Present address: Øystein Angen, Norwegian Veterinary Institute, Oslo, Norway.

***Coxiella burnetii* is responsible for Q fever, a worldwide zoonosis attributed to the inhalation of aerosols contaminated by live-stock birth products. Six draft genome sequences of European *C. burnetii* isolates from ruminants are presented here. The availability of these genomes will help in understanding the potential host specificity and pathogenicity and in identifying pertinent markers for surveillance and tracing.**

Received 17 March 2014 Accepted 25 April 2014 Published 15 May 2014

Citation Sidi-Boumedine K, Ellis RJ, Adam G, Prigent M, Angen Ø, Aspán A, Thiéry R, Rousset E. 2014. Draft genome sequences of six ruminant *Coxiella burnetii* isolates of European origin. *Genome Announc.* 2(3):e00285-14. doi:10.1128/genomeA.00285-14.

Copyright © 2014 Sidi-Boumedine et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Karim Sidi-Boumedine, Karim.SIDI-BOUMEDINE@anses.fr.

The intracellular bacterium *Coxiella burnetii* is the causative agent of Q fever, a zoonotic and an abortifacient disease which occurs worldwide. Domestic ruminants are considered to be the main source of human infection with *C. burnetii*. Indeed, it spreads from ruminants to humans via the inhalation of dust and aerosols contaminated by the birth products of livestock (1). Being mainly asymptomatic in humans and animals, the disease poses a challenge to clinicians, thus delaying diagnosis and treatment or prophylaxis.

Numerous knowledge gaps in the understanding of this organism and of Q fever epidemiology (including in domestic ruminant populations) were highlighted in a recent review (2).

We have sequenced six new *C. burnetii* strains isolated from different ruminant hosts (cattle, sheep, and goats) originating from Denmark, France, and Sweden. This corresponds to an increase in the number of available *C. burnetii* genomes originating from the main reservoir of disease (i.e., ruminants).

Each strain was grown in cell culture, and the total genomic DNA was extracted from purified *C. burnetii* and converted to sequencing libraries using the Nextera XT kits (Illumina). These were normalized and pooled before sequencing either on an Illumina GAIIx instrument with 2 × 120 paired-end reads or on an Illumina MiSeq with 2 × 250 paired-end reads. For each isolate, the A5-miseq pipeline (3) was used to perform read trimming and correction, contig assembly, crude scaffolding, misassembly correction, and final scaffolding. The scaffolds were then reordered with Mauve (4) using *C. burnetii* RSA493 (Nine Mile) as the reference genome (accession no. NC_02971). Capillary sequence data were also used to fill in some gaps in the assemblies.

Genome sequencing for the *C. burnetii* isolates Cb_B1, Cb_C2, Cb_O184, EV-Cb_C13, Cb_B18, and EV-Cb_BK10 resulted in high coverage assemblies of the ~2-Mb genomes (between 200- and 400-fold), which should represent most of the functional annotated genes and allow for comparative studies using these ge-

nomes. The final number of contigs for each strain varied between 37 (EV-Cb_BK10) and 268 (Cb_O184), with a maximum contig size of between 83,061 and 350,015 bases and N_{50} values between 17,735 and 102,549 bases.

Comparisons of the assembled genomes from this study with the completed reference genomes revealed that the majority of contig breaks in our assemblies corresponded to the positions of insertion sequence (IS) elements in the reference genomes. We were also able to identify several regions of insertions, deletions, and rearrangements. The genomes of bovine-origin isolates were most similar to the RSA493 (Nine Mile) reference genome, while the genomes of the ovine and caprine isolates were more divergent. The assemblies also indicated the presence of a single plasmid in each strain.

To provide essential foundations for effective and sustainable control or intervention strategies, it is imperative to address biodiversity, host/environmental niche correlations, and comparative strain investigations of host-microbe interactions, using refined genomic and postgenomic methods. The availability of these six new genome assemblies together with other publicly available sequence data for this species (5–8) will help in performing comparative genomics to understand the genome differences between the *C. burnetii* strains, including rearrangements, insertions, and deletions, thus contributing to the identification of relevant molecular and virulence markers.

Nucleotide sequence accession numbers. The assembled genomes of these six *C. burnetii* isolates have been deposited at the European Nucleotide Archive under accession no. [CCAH000000000](https://www.ebi.ac.uk/ena/record/CCAH000000000/), [CCAI000000000](https://www.ebi.ac.uk/ena/record/CCAI000000000/), [CCAJ000000000](https://www.ebi.ac.uk/ena/record/CCAJ000000000/), [CCAK000000000](https://www.ebi.ac.uk/ena/record/CCAK000000000/), [CCAL000000000](https://www.ebi.ac.uk/ena/record/CCAL000000000/), and [CCAM000000000](https://www.ebi.ac.uk/ena/record/CCAM000000000/).

ACKNOWLEDGMENTS

This work was supported by Collaborating Veterinary Laboratories (CoVetLab), a partnership network of national veterinary public health

institutes from Denmark, France, The Netherlands, Sweden, and the United Kingdom.

The strain Cb_B18 was isolated at INRA Nouzilly and is held by their International Center for Microbial Resources (CIRM).

We thank Annie Rodolakis for kindly providing strains Cb_B1 and Cb_C2.

REFERENCES

1. Maurin M, Raoult D. 1999. Q fever. Clin. Microbiol. Rev. 12:518–553.
2. Georgiev M, Afonso A, Neubauer H, Needham H, Thierry R, Rodolakis A, Roest H, Stark K, Stegeman J, Vellema P, van der Hoek W, More S. 2013. Q fever in humans and farm animals in four European countries, 1982 to 2010. Euro Surveill. 18:pii=20407. <http://www.eurosurveillance.org/ViewArticle.aspx?ArticleId=20407>.
3. Coil D, Guillaume J, Darling AE. 2014. A5-miseq: an updated pipeline to assemble microbial genomes from Illumina MiSeq data. Cornell University, Ithaca, NY. <http://arxiv.org/abs/1401.5130>.
4. Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One 5:e11147. <http://dx.doi.org/10.1371/journal.pone.0011147>.
5. Seshadri R, Paulsen IT, Eisen JA, Read TD, Nelson KE, Nelson WC, Ward NL, Tettelin H, Davidsen TM, Beanan MJ, Deboy RT, Daugherty SC, Brinkac LM, Madupu R, Dodson RJ, Khouri HM, Lee KH, Carty HA, Scanlan D, Heinzen RA, Thompson HA, Samuel JE, Fraser CM, Heidelberg JF. 2003. Complete genome sequence of the Q-fever pathogen *Coxiella burnetii*. Proc. Natl. Acad. Sci. U. S. A. 100:5455–5460. <http://dx.doi.org/10.1073/pnas.0931379100>.
6. Beare PA, Unsworth N, Andoh M, Voth DE, Omsland A, Gilk SD, Williams KP, Sobral BW, Kupko JJ, Porcella SF, Samuel JE, Heinzen RA. 2009. Comparative genomics reveal extensive transposon-mediated genomic plasticity and diversity among potential effector proteins within the genus *Coxiella*. Infect. Immun. 77:642–656. <http://dx.doi.org/10.1128/IAI.01141-08>.
7. Rouli L, Rolain JM, El Filali A, Robert C, Raoult D. 2012. Genome sequence of *Coxiella burnetii* 109, a doxycycline-resistant clinical isolate. J. Bacteriol. 194:6939. <http://dx.doi.org/10.1128/JB.01856-12>.
8. Karlsson E, Macellaro A, Bystrom M, Forsman M, Frangoulidis D, Janse I, Larsson P, Lindgren P, Ohrman C, van Rotterdam B, Sjödin A, Myrtenaäs K. 2014. Eight new genomes and synthetic controls increase the accessibility of rapid melt-MAMA SNP typing of *Coxiella burnetii*. PLoS One 9:e85417. <http://dx.doi.org/10.1371/journal.pone.0085417>.